

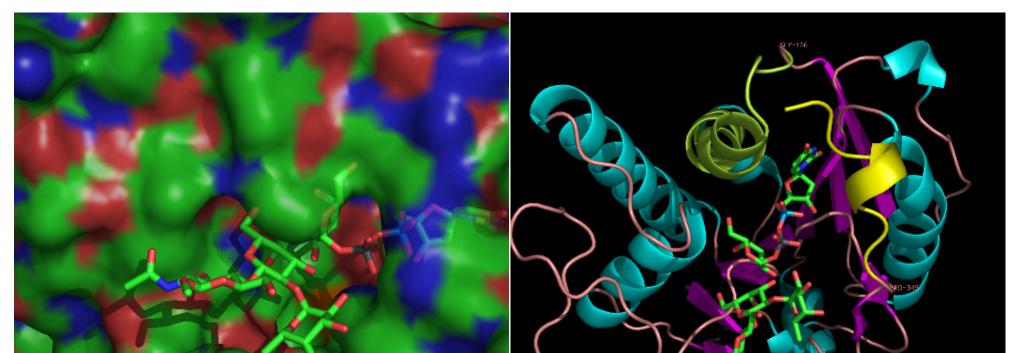
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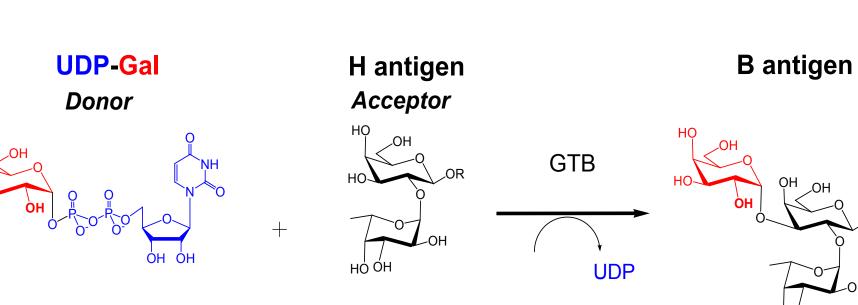
Isotope Labeling of Human Blood Group B **Galactosyltransferase for NMR Experiments**

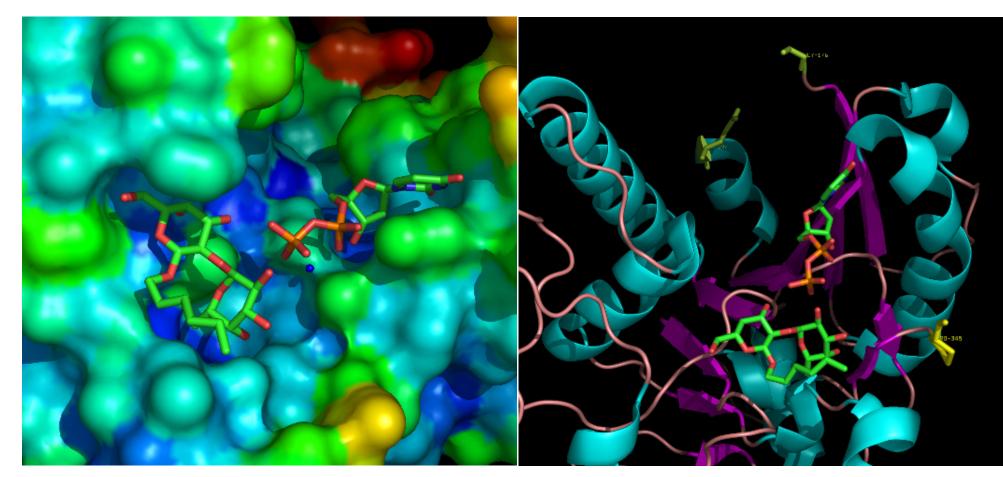
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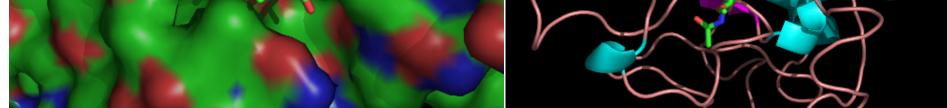
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Human Blood Group B Galactosyltransferase (GTB)









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GTB is a retaining glycosyltransferase that transfers galactose from UDP-Gal to acceptor saccharides (center). The crystal structure (right panel, [2]) does not reveal the position of the galactose residue and is missing a loop (residues 177-196) and the Cterminal end (residues 346-354). A homology model (left panel, [4]) shows that UDP-Gal should be completely buried in the donor binding pocket.

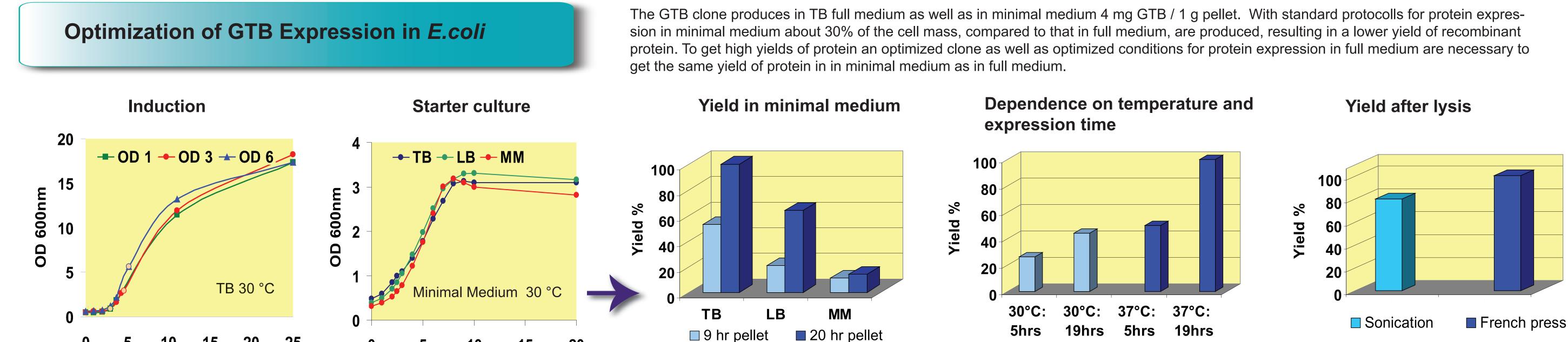
Open Questions

- What is the role of the C-terminal loop for the catalytic activity of GTB? How does the catalytic residue **Glu303 interact with the donor substrate?**
- In order to address these questions by NMR, uniformly or selectively ¹³C,²H,¹⁵N- (CDN)-isotope labeled GTB has to be used.
- Strategies for selective labeling of important amino acids in the catalytic site and in the loop are required.
- The labeling techniques have to yield high expression rates of GTB to minimize the costs.

New Achievments

Here we describe for the first time:

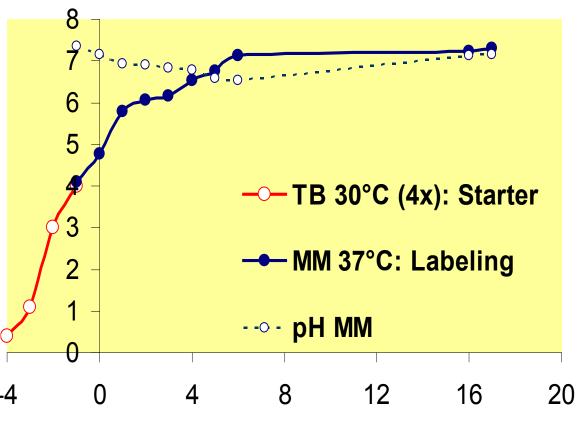
- Expression of uniformly ²H,¹⁵N-labeled GTB (DN-GTB) in high yield with a low cost labeling technique [3].
- Expression of uniformly ¹⁵N-labeled GTB (N-GTB) in high yield.
- 700 MHz NMR (cryo probe): ¹H,¹⁵N HSQC and ¹H,¹⁵N TROSY spectra of these GTB-samples.



25 20 10 15 time (hrs)

time (hrs)

- The expression on rich medium should be optimized. Expression should not depend on OD of induction.
- The yield of GTB did not decrease until induction at OD 4
- The yield of recombinant protein in minimal medium depends on the starter culture used and on the optimized supplements to the minimal medium. The growth curves in minimal medium do not show any difference although different starter cultures were used.
- 5hrs
- The temperature for expression in minimal medium can be totally different from the temperature in full medium.
- The time of expression has to be checked carefully. Some clones produce more than 60% of recombinant protein during the stationary phase.
- French Press lyses gives about 20% more yield of protein than ultrasonication.

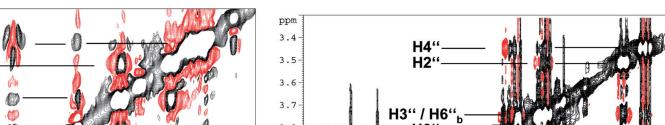


time (hours)

• For the production of DN-GTB the pellet from 500 ml TB medium at an OD of 4 is used to inoculate 500 ml DN-minimal medium (concentration factor 4x).

• The concentration of the antibiotic is doubled to increase the selection pressure in D_2O .

• Expression in minimal medium is performed at 37°C, after one hour growing the bacteria are induced. Seven hours after induction the temperature is set to 30°C and protein expression is stopped after 16 hours.



DN-minimal medium composition	final conc.	impor- tant
KD ₂ PO ₄ /Na ₂ DPO ₄	50 mM	
NaCl	0.05%	
¹⁵ ND₄CI	0.10%	
D7-glucose	0.40%	
D8-glycerol	0.40%	++
thiamin	20 µg/ml	+
MgSO ₄	4 mM	+++
MgCl ₂	1 mM	+
CaCl ₂	0.1 mM	
FeSO ₄ , CoCl ₂	1 µM	+
ZnCl ₂	0.2 mM	+
vitamin solution BME, Sigma	1x	++
DN-uniform labeled full medium	10%	++
Amp	200 µg/ml	+
рН	7.2 - 7.5	+
temperature	37°C	+++
time of expression	18 hrs	+++
air supply		++

Yield and costs of labeled GTB

Medium	ТВ	N-MM	DN-MM	Standard DN- labeling
Volume	1000 ml	1000 ml	1000 ml	1000 ml
€/L	40 €	60 €	2,800 €	2,800 €
mg / L	100 mg	100 mg	120 mg	28 mg
€ / mg	0.40€	0.60 €	23€	100 €

Labeling Strategy

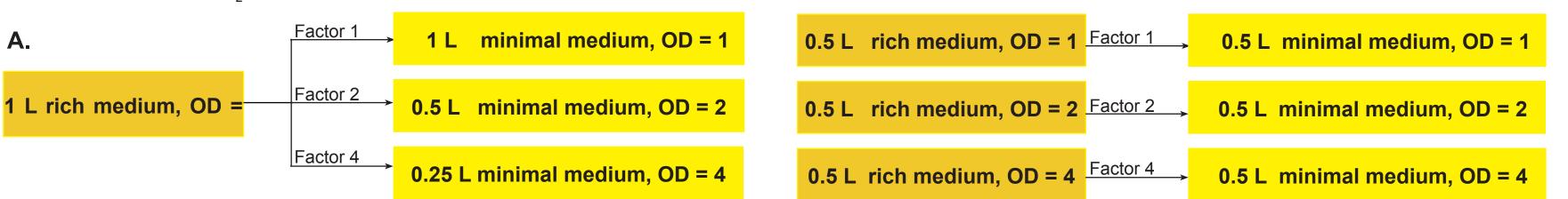
A: The labeling technique after Marley et al. [3]:

• High yields of labeled protein, significant reduction in costs for labeled compounds, isotopic incorporation >90%.

• Cell mass is generated in unlabeled rich media allowing rapid growth to high cell densities (but cells have to be still in the log-phase!).

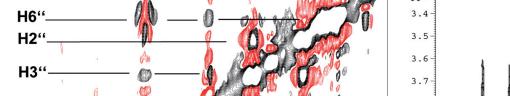
• The cell pellet from unlabeled rich medium is used to inoculate isotopically defined labeled minimal medium a higher cell densities, optimized for maximal protein expression.

• The labeling technique can be adapted to all uniform CDN-isotope labeling. No adaption of bacteria to D₂O is necessary.



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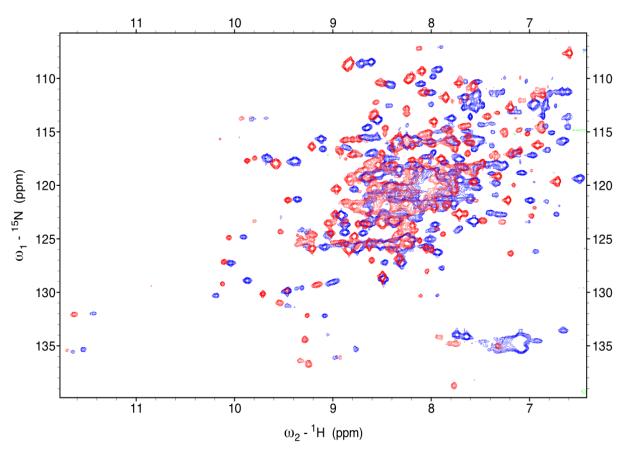
	m / p	5 -	
oncentration of	600 n	-	TB 30
volumes of rich Ition in minimal	OD 0		→ MM 3 [•]
it is possible to			

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B: Modified labeling protocoll When the concentration factor after Marley et al. is 4 and a high co the labeled protein is needed, it can be difficult to manage the high v

medium with an OD of 1 to produce the cell mass needed for inoculat medium.

• If the clone accepts an induction at OD 4 without any loss in yield it is possible to inoculate the same volume of minimal medium with the bacterial pellet at an OD

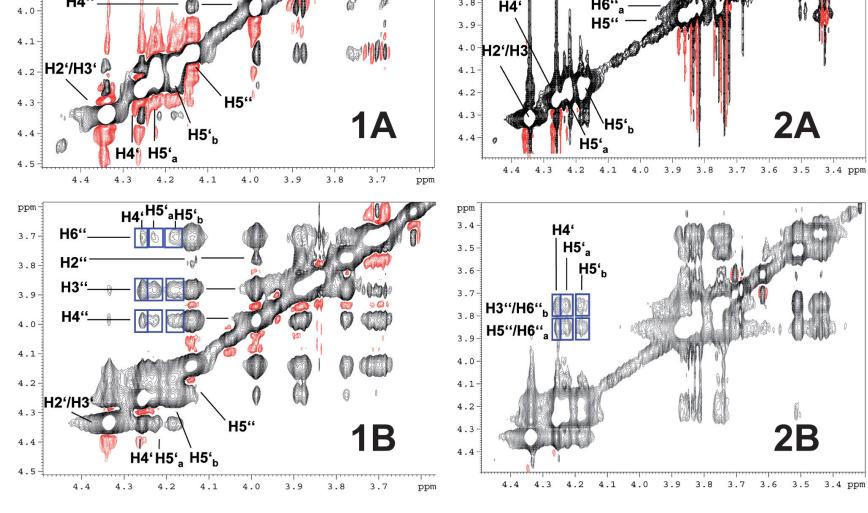


¹H,¹⁵N TROSY spectra at 700 MHz of 800 µM DN-GTB in 25 mM BisTris pH 6.5.* GTB contains only 20% α -helix and 20% β -sheet. Red: 50°C, blue: 25°C.

we used perdeuterated DN-GTB for the transferred NOE experiments. From MALDI-TOF spectra we estimated the extent of deuteration to be larger than 95%. • The NOESY spectra obtained for UDP-Gal and UDP-Glc in the presence of perdeuterated DN-GTB at 700 MHz unambiguously show negative NOE cross peaks. • It is well documented that bivalent cations such as Mg²⁺ and Mn²⁺ have a significant impact on the function of glycosyltransferases. With NMR we have directly observed the conformational transitions that occur upon binding of the donor ligands, UDP-Gal and UDP-Glc to GTB in the presence of Mg^{2+} [5]. • NOEs across the pyrophosphate bridge (blue squares)

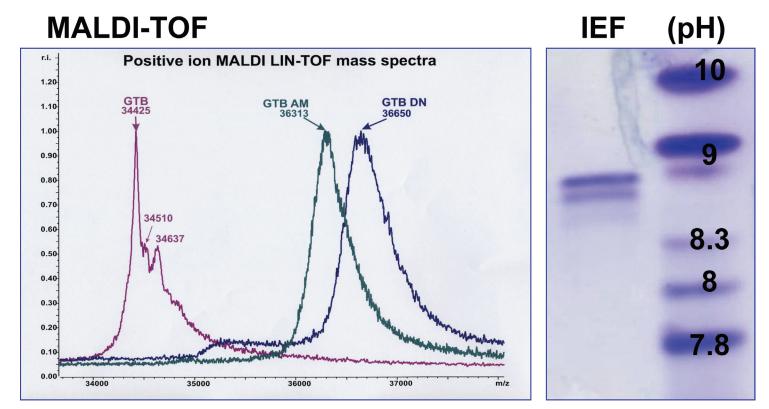
are only observed for the bound states of UDP-Gal and UDP-Glc.

• In the absence of Mg²⁺ ions UDP-Gal and UDP-Glc are bound to GTB but no NOEs across the pyrophosphate bridge are observed.



¹H-NOESY spectra of a complex of DN-GTB with UDP-Gal (1) and UDP-Glc (2) in the absence (A) and in the presence (B) of Mg^{2+} ions in 25 mM BisTris, pH 6.5* at 700 MHz. The protein : ligand ratio is 1:2.





The incorporation level of DN-GTB is >90% as determined with MALDI-TOF. Labeled and unlabeled GTB show two isoforms in the isoelectrofocussing gel.

* 25 mM BisTris, pH 6.5 (RT), 50 mM NaCl, 10 mM MgCl₂, 5 mM 2ME

References:

- [1] M.M. Palcic, N.O.L. Seto, O. Hindsgaul (2001) Transf. Med. 11, 315-323
- [2] S.I.Patenaude, N.O.L. Seto, S.N. Borisova, A. Szpacenko, S.L. Marcus, M.M. Palcic, S.V. Evans (2002) Nature Struct. Biol., 9, 685-690

[3] J. Marley, M. Lu, C. Bracken (**2001**) *J. Biomol. NMR*, 20, 71-75;

- [4] Heissigerová H.; Breton C.; Moravcová J.; Imberty A., *Glycobiology*, 2003, 13, 377-386
- [5] J. Angulo, B. Langpap, A. Blume, T. Biet, B. Meyer, N.R. Krishna, H. Peters, M.M. Palcic, T. Peters (2006), submitted

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